

qRT-PCR Protocol

Reagents/Notes

- Use Luna® Universal Probe One-Step RT-qPCR Kit (NEB, E3006L)
- Use IDT Tool (<https://www.idtdna.com/scitools/Applications/RealTimePCR/Default.aspx>) to design primers and probes—desired amplicon size: 75-200 bp
- Primers => resuspend to 200 uM in RNase-free ddH2O
- Probes => 5' 6-FAM, 3' TAMRA, HPLC purified, resuspend to 100 uM in RNase-free ddH2O
- Use RNase free technique: separate filter tips, spray work area down with RNase Zap, RNase-free ddH2O

1. Isolate RNA—See RNA Isolation Protocol
 - a. Run Taq PCR to determine if DNA contamination is present in RNA
 - i. Can use any PCR amplicon that amplifies easily
 - b. If no amplification present—continue with RT-PCR protocol
2. Nanodrop RNA
 - a. Dilute RNA samples to 5 ng/ul in RNase-free ddH2O
3. Make RNA standard curve
 - a. Start with 5 ng/ul WT control RNA (1×10^{-9})
 - b. Make 4 successive 1:1 dilutions in RNase-free ddH2O
 - c. Final concentrations: 1×10^{-9} , 5×10^{-10} , 2.5×10^{-10} , 1.25×10^{-10} , 6.25×10^{-11}
4. Set up qRT-PCR Mastermix (on ice!)
 - a. Always include wells for Act1 control
 - b. Construct separate mastermix for each primer/probe combination:

Reaction Mixture (18 ul)

2x One-Step Reaction mix:	10 ul
2x Warmstart RT mix:	1 ul
Primer 1 (20 uM):	0.4 ul
Primer 2 (20 uM):	0.4 ul
Probe (10 uM):	0.4 ul
RNase-free ddH2O:	5.8 ul

5. Aliquot Mastermix (in chilled PCR rack)—see plate layout on next page
 - a. 18 ul of mastermix per well (in triplicate)
6. Add 2 uL of 5 ng/ul RNA to corresponding wells (10 ng total RNA/reaction)

